Preparation of a (±)-1,6-Di-O-feruloyl-myo-inositol **Derivative: An Efficient Method for** Introduction of Ferulic Acid to 1,6-Vicinal Hydroxyl Groups of *myo*-Inositol

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Ferulic acid (1) can be synthesized by the reaction of vanillin with malonic acid in the presence of an amine.¹ However, this reaction requires a long reaction time for completion and, hence, is impractical as a method for a large scale preparation of 1.¹ On the other hand, 1 is known to occur in large amounts in various renewable resources such as rice, wheat, and corn. Recently, we have developed an efficient and practical method for the mass production of 1 from the oily component of rice bran.² Since then, the synthetic applications of 1 and related compounds have attracted considerable attention, especially in the field of chemoprevention study. For example, the ferulic acid derivative, ethyl 3-(4'-geranyloxy-3'-methoxyphenyl)-2-propenoate,^{2b} in which the geranyl group is attached to the phenolic hydroxyl group of ethyl ferulate, was shown to have a suppressive effect on the formation of a colonic tumor marker in rats.³ O-Tocopheryl succinyl O-ethyl ferulate was found to reduce the replication rate of the HIV-1 virus in infected cells in vitro by 80%.⁴

myo-Inositol is also obtained from rice bran, and it plays an important role in biological systems. D-mvo-Inositol-1,4,5-trisphosphate was found to act as an intracellular second messenger for calcium mobilization.^{5,6} myo-Inositol hexaphosphate (IP₆) was shown to have an anticancer action in a variety of experimental tumor models.7

Ferulic acid occurs usually as various ester or amide derivatives in natural products,⁸ but the ester compounds containing ferulic acid and *myo*-inositol as components have not yet been discovered in nature.



In a previous paper, we reported the synthesis of seven ester compounds consisting of ferulic acid and myoinositol and their inhibitory effect on 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced superoxide (O_2^{-}) generation by the use of differentiated HL-60 cells.⁹ In this study, we found that only 3,4,5,6-tetra-O-acetyl-1,2di-O-[3-(4'-acetoxy-3'-methoxyphenyl)-2-propenoyl]-myoinositol (2) showed a distinct inhibitory activity. This result suggests that the architecture having two feruloyl groups at the vicinal positions of myo-inositol is essential for the inhibitory activity.

In the present paper, we report an efficient method for the introduction of ferulic acid to the 1,6-vicinal hydroxyl groups of (\pm) -3,4-O-(1,1,3,3-tetraisopropyldisiloxanedi-1,3-yl)-myo-inositol (TIPDS-myo-inositol, 4) and the effect of intermolecular hydrogen bonding of 4 on the esterification reaction.

The introduction of functional groups to the 1,6-vicinal hydroxyl groups of myo-inositol has been performed through 2,3:4,5-di-O-cyclohexlidene-myo-inositol as an intermediate.¹⁰ However, the synthesis of this diketal intermediate was difficult and troublesome: the reaction of myo-inositol with cyclohexanone gave 1,2-O-cyclohexylidene-myo-inositol and three diketals (1,2:3,4-, 1,2:4,5-, and 2,3:4,5-di-O-cyclohexylidene-myo-inositol).11 The reaction of myo-inositol with 1-ethoxycyclohexene in DMF also gave the three diketals.¹² In these reactions, the yield of the 2,3:4,5-diketal was low, and the isolation of this ester from the reaction mixture was tedious.

In contrast, the disiloxanyl derivative **3** can readily be obtained from *myo*-inositol by consecutive regioselective cyclohexylidenation and silvlation reactions.^{13,14} The reaction of **3** with 1 equiv of ethylene glycol in the presence of a catalytic amount of *p*-toluenesulfonic acid in CHCl₃ at room temperature gave **4** as a white powder in an 86% yield (Scheme 1). We then examined the introduction of ferulic acid to the 1,6-vicinal hydroxyl groups of 4.

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 a Reagents and conditions: $Et_3N,\ DMAP,\ and\ CH_2Cl_2,\ room temperature, 12 h.$



Figure 1. IR absorption spectra of **4** in CH_2Cl_2 : (a) 2.5 mM, (b) 10 mM, and (c) 50 mM.

Ferulic acid was converted into 3-(4'-acetoxy-3'-methoxyphenyl)-2-propenoyl chloride (5) via two steps. The reaction of **4** with **5** was carried out in the presence of a mixture of triethylamine and 4-(dimethylamino)pyridine (DMAP) in CH_2Cl_2 (Scheme 2). However, the yield of the desired ester was very low.

The acid chloride **5** usually has sufficient reactivity in the ester formation with an alcohol.⁹ Therefore, we studied the reason for the poor reactivity of **4** in this reaction. The solubility experiment revealed the unexpected fact that **4** was soluble in hydrophobic solvents. This fact implies that intramolecular and/or intermolecular hydrogen bonding exists in **4** in hydrophobic solvents such as CH_2Cl_2 .

The FT-IR spectra of **4** in a 50 mM CH_2Cl_2 solution showed three peaks at 3688, 3585, and 3450 cm⁻¹ in the OH-stretching region (Figure 1). The absorption peaks at 3688 and 3585 cm⁻¹ can be assigned to the free OH group and the intramolecularly hydrogen-bonded OH group, respectively.¹⁵⁻¹⁷ The absorption band centered at



Figure 2. $^1\mathrm{H}$ NMR spectra (400 MHz) of 4 in $\mathrm{CD}_2\mathrm{Cl}_2$ at various concentrations.

3450 cm⁻¹ can be assigned to the intermolecularly hydrogen-bonded OH group, because the absorbance of the band increased with increases in the concentration of **4**.

The formation of intermolecular hydrogen bonding was also confirmed by measuring the ¹H NMR spectrum of **4** in CD_2Cl_2 at various concentrations (Figure 2). The OH proton signals appeared at 2.4–2.7 ppm in the 2.5 mM solution, and these signals shifted downfield with increases in the concentration of **4**. These results strongly suggest that the OH groups of **4** form intermolecular hydrogen bonds in the real reaction mixture, and this intermolecular hydrogen bonding results in the poor reactivity in the esterification reaction of **4** with **5** in CH₂-Cl₂ solution.

In general, intermolecular hydrogen bonding can be dissociated by raising the temperature or by the addition of a cosolvent such as DMF, THF, or pyridine; in fact, it is well-known that the esterification of an acid chloride by an alcohol proceeds efficiently in pyridine.¹⁸

We also found that the reaction of **4** with **5** in pyridine proceeded smoothly, and the ester compounds **6a**-**d** were obtained in moderate yields. When the reaction was carried out in pyridine in the presence of DMAP at 70 °C using **4** and **5** in a molar ratio of 1:5, **6c** was selectively obtained in an 81% yield. This regioselectivity is also compatible with the facts that (1) the hydroxyl group at the 2-position of **4** has a poor reactivity due to a hindered axial bond¹⁹ and (2) the hydroxyl group at the 5-position of **4** is protected by bulky silyl substituents.^{14,20}

Experimental Section

General Methods. The ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra were recorded on a Varian unity-plus 400 spectrometer. FT-IR spectra were recorded on a Shimazu FTIR 8200D instrument. The solutions were measured between NaCl windows. Solid samples were measured as KBr pellets using a JASCO IR-810 instrument. Melting points were measured in capillary tubes with a Yanagimoto micro melting point apparatus and are uncorrected. Elemental analyses were performed on a Perkin-Elmer 2400II instrument. Compound purity

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was checked by TLC on Silica Gel 60 F254 (E. Merck) with detection by charring with phosphomolybdic acid (10% in EtOH solution). Column chromatography was performed on Silica Gel, Wakogel C-200 (Wako Pure Chemical Industry).

Materials. Ferulic acid and *myo*-inositol were provided by Tsuno Food Industrial Co., Ltd. Ferulic acid was recrystallized from ethanol. *myo*-Inositol was dried under reduced pressure in a drying oven. (\pm) -1,2-*O*-Cyclohexylidene-3,4-*O*-(1,1,3,3-tetraisopropyldisiloxanedi-1,3-yl)-*myo*-inositol (**3**) was prepared according to literature.^{13,14} Other chemicals were commercial products and were used without further purification. Solvents were reagent grade and, in most cases, dried prior to use.

(±)-3,4-*O*-(1,1,3,3-Tetraisopropyldisiloxanedi-1,3-yl)-*myo* inositol (4). A mixture of a catalytic amount of *p*-toluenesulfonic acid and ethylene glycol (0.74 g, 12.0 mmol) was added to a solution of **3** (5.48 g, 10.9 mmol) in CHCl₃ (50 mL). The mixture was stirred at room temperature for 1 h. The organic layer was washed with saturated aqueous NaHCO₃ and water, and dried over Na₂SO₄. The solvent was evaporated to dryness under reduced pressure by an aspirator. The residue was chromatographed (silica gel, CHCl₃) to give **4** as a white solid (3.97 g, 86%): mp 118–121 °C; ¹H NMR (DMSO-*d*₆) δ 4.45–4.65 (m, 4H), 3.68–3.75 (m, 2H), 3.51 (dd, 1H, *J* = 2.4, 9.2 Hz), 3.39 (m, 1H), 3.19 (m, 1H), 3.02 (m, 1H), 0.86–1.10 (m, 28H); ¹³C NMR (DMSO-*d*₆) δ 77.00, 75.61, 75.08, 72.97, 72.38, 71.51, 17.61, 17.57, 17.50, 17.39, 17.22, 12.54, 12.00, 11.87. Anal. Calcd for C₁₈H₃₈O₇Si₂: C, 51.15; H, 9.06. Found: C, 51.12; H, 9.21.

3-(4'-Acetyloxy-3'-methoxyphenyl)-2-propenoic Acid Chloride (5). Ferulic acid (1) (38.8 g, 0.2 mol) was added to a solution of NaOH (25.4 g, 0.52 mol) in water (200 mL), and the mixture was cooled to below 10 °C. Acetic anhydride (25.4 g, 0.25 mol) was then added to the cold solution. The solution was stirred at 20 °C for 10 min and then at room temperature for 20 min. Then, the pH of the solution was adjusted to 4–5 by adding dilute sulfuric acid. The resulting white precipitate was filtered and washed with water. Recrystallization from EtOH gave colorless needles (37.7 g, 77%): mp 197–200 °C; ¹H NMR (DMSO-*d*₆) δ 12.45 (s, 1H), 7.61 (d, 1H, *J* = 15.6 Hz), 7.12–7.49 (m, 3H), 6.61 (d, 1H, *J* = 15.6 Hz), 3.84 (s, 3H), 2.28 (s, 3H); ¹³C NMR (DMSO-*d*₆) δ 168.6, 167.8, 151.3, 143.5, 141.0, 133.4, 123.4, 121.5, 119.7, 112.0, 56.2, 20.6.

Thionyl chloride (8.5 mL, 0.12 mol) was added to a suspension of 3-(4'-acetyloxy-3'-methoxyphenyl)-2-propenoic acid (23.6 g, 0.1 mol) in CHCl₃ (100 mL) containing a catalytic amount of DMF. The mixture was stirred for 4 h at the reflux temperature. After the solvent and excess thionyl chloride were removed under reduced pressure, the acid chloride **5** was obtained as pale yellow crystals (25.0 g, 98%). This material was used without further purification: mp 129–131 °C; ¹H NMR (CDCl₃) δ 7.79 (d, 1H, *J* = 15.6 Hz), 7.09–7.12 (m, 3H), 6.59 (d, 1H, *J* = 15.6 Hz), 3.88 (s, 3H), 2.35 (s, 3H); ¹³C NMR (CDCl₃) δ 168.5, 165.9, 151.7, 149.8, 142.8, 131.8, 123.6, 122.5, 122.4, 111.9, 56.0, 20.6. Anal. Calcd for C₁₂H₁₁ClO₄: C, 56.60; H, 4.35. Found: C, 56.56; H, 4.28.

(\pm)-1-*O*-[3-(4'-Acetyloxy-3'-methoxyphenyl)-2-propenoyl]-3,4-*O*-(1,1,3,3-tetraisopropyldisiloxanedi-1,3-yl)-*myo*-inositol (6a) and (\pm)-6-*O*-[3-(4'-Acetyloxy-3'-methoxyphenyl)-2propenoyl]-3,4-*O*-(1,1,3,3-tetraisopropyldisiloxanedi-1,3yl)-*myo*-inositol (6b). The acid chloride 5 (159 mg, 0.625 mmol) was added to a solution of 4 (106 mg, 0.25 mmol) in anhydrous pyridine (10 mL). The mixture was stirred for 14 h at room temperature, and the reaction was quenched by adding water. Ethyl acetate was added, and the mixture was washed successively with saturated aqueous KHSO₄, saturated aqueous NaHCO₃, and brine. After the organic layer was dried over Na₂-SO₄, the solvent was removed under reduced pressure. The residue was chromatographed on preparative TLC (1 mm thickness of silica gel, 3:2 *n*-hexane/ethyl acetate) to give **6a** (52.8 mg, 33%) and **6b** (10.5 mg, 6.6%) as white solids, respectively. **6a:** TLC (1:1 *n*-hexane/ethyl acetate) $R_f = 0.35$; mp 163–165 °C; ¹H NMR (CDCl₃) δ 7.72 (d, 1H, J = 16.0 Hz), 7.04–7.13 (m, 3H), 6.54 (d, 1H, J = 16.0 Hz), 4.97 (dd, 1H, J = 2.8, 10.0 Hz), 4.25 (t, 1H, J = 2.8 Hz), 4.18 (m, 1H), 3.95 (t, 1H, J = 8.8 Hz), 3.86 (s, 3H), 3.79 (dd, 1H, J = 2.8, 8.8 Hz), 3.45 (m, 1H), 2.59–2.62 (m, 2H), 2.51 (d, 1H, J = 3.2 Hz), 2.33 (s, 3H), 1.00–1.11 (m, 28H); ¹³C NMR (CDCl₃) δ 168.76, 166.51, 151.36, 145.23, 141.57, 133.17, 123.24, 121.61, 117.60, 111.08, 76.15, 75.06, 74.76, 73.06, 71.00, 69.59, 55.88, 20.65, 17.44, 17.30, 17.20, 17.12, 12.81, 12.76, 12.06, 12.03. Anal. Calcd for C₃₀H₄₈O₁₁Si₂: C, 56.22; H, 7.55. Found: C, 56.19; H, 7.60.

6b: TLC (1:1 *n*-hexane/ethyl acetate) $R_f = 0.49$; mp 102–104 °C; ¹H NMR (CDCl₃) δ 7.71 (d, 1H, J = 16.0 Hz), 7.04–7.13 (m, 3H), 6.47 (d, 1H, J = 16.0 Hz), 5.40 (t, 1H, J = 10.0 Hz), 4.14 (t, 1H, J = 2.8 Hz), 3.98 (t, 1H, J = 8.8 Hz), 3.86 (s, 3H), 3.53–3.73 (m, 3H), 2.70–2.78 (m, 2H), 2.45 (d, 1H, J = 3.2 Hz), 2.33 (s, 3H), 1.00–1.11 (m, 28H); ¹³C NMR (CDCl₃) δ 168.76, 167.18, 151.32, 144.92, 141.42, 133.35, 123.21, 121.36, 117.80, 111.23, 74.67, 74.10, 73.06, 72.77, 70.92, 55.88, 20.66, 17.41, 17.32, 17.27, 17.20, 17.11, 12.78, 12.05, 12.02. Anal. Calcd for C₃₀H₄₈O₁₁Si₂: C, 56.22; H, 7.55. Found: C, 56.17; H, 7.63.

(±)-1,6-O-Bis[3-(4'-acetyloxy-3'-methoxyphenyl)-2-propenoyl]-3,4-O-(1,1,3,3-tetraisopropyldisiloxanedi-1,3-yl)-myoinositol (6c) and (±)-1,5,6-O-Tris[3-(4'-acetyloxy-3'-methoxyphenyl)-2-propenoyl]-3,4-O-(1,1,3,3-tetraisopropyldisiloxanedi-1,3-yl)-myo-inositol (6d). A mixture of a catalytic amount of 4-(dimethylamino)pyridine and 5 (318 mg, 1.25 mmol) was added to a solution of 4 (106 mg, 0.25 mmol) in anhydrous pyridine (10 mL). The mixture was stirred at 70 °C for 1.5 h, and the reaction was quenched by adding water. Ethyl acetate was added, and the mixture was washed successively with saturated aqueous KHSO₄, saturated aqueous NaHCO₃, and brine. After the organic layer was dried over Na₂SO₄, the solvent was removed under reduced pressure. The residue was chromatographed (silica gel, 3:2 *n*-hexane/ethyl acetate) to give **6c** as a white solid (174 mg, 81%): TLC (1:1 *n*-hexane/ethyl acetate) $R_f = 0.57$; mp 198–200 °C; ¹H NMR (CDCl₃) δ 7.57–7.63 (m, 2H), 6.97-7.04 (m, 6H), 6.30-6.40 (m, 2H), 5.72 (t, 1H, J=10.0 Hz), 5.35-5.38 (m, 2H), 4.99 (dd, 1H, J=2.4, 10.0 Hz), 4.06 (m, 1H), 3.91 (m, 1H), 3.77-3.84 (m, 7H), 3.55 (m, 1H), 2.24 (s, 6H), 0.85-1.10 (m, 28H); ¹³C NMR (CDCl₃) & 168.69, 166.18, 166.12, 151.37, 151.31, 145.48, 144.95, 141.64, 141.48, 133.20, 133.02, 123.17, 123.20, 121.76, 121.45, 117.59, 117.21, 111.17, 111.06, 74.61, 73.41, 71.10, 70.97, 70.80, 55.92, 55.87, 20.61, 17.46, 17.35, 17.32, 17.26, 17.21, 17.13, 12.79, 12.06. Anal. Calcd for C42H58O15-Si₂: C, 58.72; H, 6.81. Found: C, 58.67; H, 6.94.

Further elution gave **6d** as a white solid (29.6 mg, 11%): TLC (1:1 *n*-hexane/ethyl acetate) $R_f = 0.47$; mp 191–194 °C; ¹H NMR (CDCl₃) δ 7.52–7.67 (m, 3H), 7.00–7.08 (m, 9H), 6.24–6.43 (m, 3H), 5.91 (t, 1H, J = 10.4 Hz), 5.37 (t, 1H, J = 10.0 Hz), 5.27 (dd, 1H, J = 2.4, 10.0 Hz), 4.33 (t, 1H, J = 2.8 Hz), 4.26 (t, 1H, J = 9.2 Hz), 3.95 (dd, 1H, J = 2.8, 8.4 Hz), 3.84 (s, 6H), 3.81 (s, 3H), 2.31 (s, 6H), 2.29 (s, 3H), 0.90–1.12 (m, 28H); ¹³C NMR (CDCl₃) δ 168.77, 168.68, 166.00, 165.77, 165.53, 151.36, 151.30, 145.61, 145.19, 144.93, 141.67, 141.54, 141.49, 133.21, 133.03, 123.21, 123.15, 121.80, 121.64, 121.39, 117.36, 117.12, 111.17, 111.09, 75.12, 74.27, 72.74, 71.00, 70.89, 69.22, 55.94, 55.91, 20.64, 17.45, 17.34, 17.24, 17.14, 17.08, 12.79, 12.09, 12.03. Anal. Calcd for C₅₄H₆₈O₁₉Si₂: C, 60.21; H, 6.36. Found: C, 60.18; H, 6.44.

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